Dendritic cells: inciting and inhibiting autoimmunity Shannon J Turley

Dendritic cells are considered the most influential antigen presenting cells in the body because of their unique role in initiating immunity against threatening antigens. Recent studies addressing the consequences of self-antigen presentation by dendritic cells revealed the unexpected ability of these antigen presenting cells to inhibit T cell-mediated autoimmune diseases. The specific mechanisms by which dendritic cells suppress immune responses have been explored during the past year. These efforts indicate that extrathymic dendritic cells control autoimmunity by inducing peripheral T cell tolerance, a function intimately linked to their state of maturation.

Address

Department of Immunology and Immunogenetics, Joslin Diabetes Center, Harvard University Medical School, One Joslin Place, Boston, MA 02215, USA

Current Opinion in Immunology 2002, 14:765-770

0952-7915/02/\$ - see front matter © 2002 Elsevier Science Ltd. All rights reserved.

Published online 4 October 2002

Abbreviations

APC	antigen presenting cell
DC	dendritic cell
EAE	experimental autoimmune/allergic encephalomyelitis
IFN	interferon
IL	interleukin
LCMV	lymphocytic choriomeningitis virus
LN	lymph node
RA	rheumatoid arthritis
SLE	systemic lupus erythematosus
Th	Thelper
	-

Introduction

Dendritic cells (DCs) are bone marrow-derived antigen presenting cells (APCs), unrivalled in their capacity for activating naïve and effector T cells. The life history of DCs unfolds in two main developmental stages, termed immature and mature. Immature DCs form lattice-like networks in virtually every tissue where they peruse the extracellular milieu, avidly endocytosing diverse antigens. Signaling by select pathogens, pro-inflammatory mediators, or CD40L, triggers immature DCs to embark on an irreversible differentiation process that results in mature DCs displaying remarkable immunostimulatory might. Before reaching peak maturity, DCs pass through an intermediate stage that is obligatory for Langerhans cells (LCs), red pulp splenic DCs and bone marrow-derived DCs. At this stage, DCs exhibit a transitional or semi-mature phenotype in terms of TCR ligand and accessory molecule expression; however, their functional contributions have not been well characterized. Maturation radically boosts the immunogenicity of DCs by inducing the stable expression of peptide-MHC complexes, upregulation of costimulatory and adhesion molecules, secretion of chemokines and

stimulatory cytokines, and swift migration to T cell zones of regional lymph nodes (LNs). As mature DCs are poised for the optimal stimulation of naïve T lymphocytes [1], antigen presentation by mature DCs is a critical checkpoint in the generation of primary immune responses.

Central tolerance is an imperfect process, thereby allowing some autoreactive T lymphocytes to escape riddance in the thymus. DCs undoubtedly process self-proteins that are either expressed endogenously or acquired during endocytosis. DC presentation of self-antigens during infection or tissue injury could lead to the misguided generation of autoaggressive T lymphocytes. Despite the presence of autoreactive lymphocytes in the circulation and the presentation of self-epitopes by 'nature's adjuvant', most individuals escape the pathological consequences of autoimmunity. Thus, extrathymic mechanisms for subduing the autoreactive lymphocyte repertoire must exist. These elusive mechanisms are collectively referred to as peripheral tolerance. Emerging evidence indicates that DCs are responsible for the establishment of peripheral tolerance as well as immunity [2]. Genetic or environmental factors that alter the immunostimulatory capacity of DCs could impair peripheral tolerance induction leading to the onset of autoimmune disease.

The paradoxical ability of DCs to incite and inhibit autoimmune disease, as well as their features in autoimmune tissues, are reviewed here. Recent studies examining the specific mechanisms by which DCs induce peripheral tolerance are also discussed.

Dendritic cells provoke and prevent autoimmune disease

DCs are the only professional APCs capable of provoking autoimmune disease to date. The transfer of DCs, isolated from donors with acute autoimmune disease or propagated in vitro under conditions that induce maturation, generates a strong T helper (Th)-1 response, eventually culminating in autoimmune disease [3-7]. This ability is restricted to DCs that have been exposed to potent maturation stimuli in the presence of abundant self-antigen. In addition, chronic maturation of tissue DCs within their native microenvironment can induce severe organ-specific autoimmune disease and systemic autoimmunity [8,9]. Therefore, presentation of self-antigens by DCs leads to autoimmune pathogenesis when the immunogenicity of the DCs is amplified. Several recent studies have established that DCs also inhibit autoimmune disease. The protection elicited by DCs is generally long lasting, antigen dependent and transferable, suggesting that DCs prevent autoimmunity by actively inducing peripheral tolerance [10,11•,12–15]. If DCs are essential for establishing tolerance to peripheral antigens, then one might speculate that autoimmune disease

results from defects in this function of DCs. In examining the mechanisms of peripheral tolerance induction by DCs, we might begin to understand how autoimmune disease unfolds.

Controlling autoimmunity by peripheral tolerance induction

Elimination of autoreactive T lymphocytes

How do the same APCs that mount primary immune responses and precipitate autoimmunity also inhibit autoimmune disease? One possibility is that tolerance is mediated by immature or semi-mature DCs expressing low levels of T cell-receptor ligands and costimulatory molecules, whereas immunity is generated by mature DCs expressing high levels of these molecules. This would require the presentation of tissue antigens by immature DCs in secondary lymphoid tissue, a scenario that seems to conflict with dogma at first glance; however, this idea may not be at odds with the classical notion of 'DC migration upon maturation' because some DCs migrate on a continuous basis [16]. Homing of migratory DCs to secondary lymphoid tissue, in the absence of maturation stimuli, could allow immature or semi-mature DCs to present tissue antigens to cognate T cells in a substimulatory context. But do DCs actually traffic peripheral antigens to regional LNs in the steady state? For some time it has been known that transport of antigenic cargo is critical for generating immunity against pathogens that invade non-lymphoid tissues.

Administration of trackable antigens, such as microbes, contact sensitisers or latex microparticles, revealed that transport of antigen from non-lymphoid to lymphoid tissue is facilitated by DCs. Because such procedures inevitably release pro-inflammatory mediators, they cannot be used to assess antigen delivery in the steady state. This dilemma was recently resolved by two studies in which gut- and skin-restricted antigens were identified within DCs of nearby LNs [17,18[•]]. Indeed, a population of migratory DCs samples antigens in peripheral tissues and transports them to draining LNs under homeostatic conditions.

The role of immature DCs in T cell engagement has largely been ignored for two main reasons: first, immature DCs are inefficient at antigen processing; and second, their positioning in peripheral tissues makes an encounter with naïve T cells unlikely. That DCs transport peripheral antigens to LNs in the steady state, however, implies that this process is carried out by immature DCs, and that antigen presentation by these cells has immunological consequences. In an important investigation, Hawiger et al. [19**] addressed these issues and found that presentation of antigen by DCs in the absence of inflammation or infection leads to *bona fide* tolerance. Soluble antigen was targeted to the MHC class II pathway of DCs in situ by non-inflammatory measures, using an antibody specific for a specialized DC endocytosis receptor. Under these conditions, antigen presentation by DCs prompted a short proliferative burst of cognate CD4+ T cells followed by their deletion. The lack of a response to subsequent peptide immunization indicated that the recipients had been rendered

tolerant. In agreement with a previous report that links peripheral tolerance to bone marrow-derived APCs, the Hawiger study established that DCs induce peripheral tolerance by eliminating autoreactive T cells [20]. In contrast, when antigen targeting was accompanied by a strong DC maturation stimulus, such as anti-CD40, the outcome was converted to immunity. These findings indicate that selfantigen presentation by immature DCs is pivotal in the elimination of autoreactive T lymphocytes and that the fate of any immune response is shaped by the 'maturity' of the DC presenting antigen.

Mature or semi-mature DCs may also contribute to the maintenance of peripheral tolerance by deleting specific subsets of autoreactive T cells. In an in vitro system described by Albert et al. [21•], memory CD8+ T cells are tolerized by DCs matured in TNF- α and prostaglandin E2 (PGE2), but not by macrophages or immature DCs. Maturation triggered by CD40 signaling, however, changed the outcome from T cell elimination to cytotoxic T lymphocyte (CTL) generation. Thus, the combination of TNF- α and PGE2 conditioned the DCs differently than anti-CD40 treatment, possibly by providing only partial maturation stimuli or by activating entirely distinct signal transduction pathways that render the DC tolerogenic. Whatever the case may be, some degree of DC maturation may be necessary for peripheral tolerance induction. It has been hypothesized that autoimmune syndromes result from defects in peripheral tolerance induction. If mature DCs participate in peripheral tolerance, then genetic or environmental factors that impair DC maturation could hinder the suppression of autoreactive T lymphocytes resulting in unchecked activation of these cells. Interestingly, blood DCs obtained from patients with autoimmune diabetes, systemic lupus erythematosus (SLE), Grave's disease and multiple sclerosis (MS), as well as individuals at risk of diabetes, exhibit a relatively immature phenotype [22-26]. When compared with healthy controls, DCs derived from patient blood were impaired in T cell stimulation and generally expressed lower levels of costimulatory molecules. Alternatively, semi-mature DCs may be required for the induction of regulatory cells, suggesting that functionally impaired DCs underlie immunoregulatory defects in autoimmune patients.

Finally, peripheral tolerance might be mediated by a specialized subset of DCs in secondary lymphoid tissue that constitutively expresses and presents peripheral antigens for the purpose of perpetual elimination of autoreactive T cells. Pugliese *et al.* [27[•]] recently identified a small subset of spleen DCs expressing pancreatic islet-specific antigens. Intriguingly, apoptotic lymphocytes were found in close proximity to DCs expressing self-antigen, suggesting that this DC population may induce T cell tolerance by direct induction of programmed cell death.

Altering the effector functions of T cells

DCs also induce peripheral tolerance by generating regulatory T cells that impede the functions of effector T cells through suppressive cytokines or a contact-dependent mechanism [28]. IL-10-producing CD4+ and CD8+ regulatory T cells can be induced by immature DCs [29,30••,31••]. It has been proposed that peripheral tolerance is maintained by immature DCs presenting antigens that are captured during normal cellular turnover, although no convincing link has been made between the presentation of apoptotic material and the induction of regulatory T cells by DCs [32]. A role for this form of tolerance induction was recently investigated in a model of autoimmune diabetes. Here, the induction of regulatory T cells by immature DCs correlated with disease prevention [33**]. Importantly, protection from diabetes appeared to be dependent on presentation of antigen derived from apoptotic β cells. DCs at an intermediate stage of maturation are also equipped to inhibit experimental autoimmune encephalomyelitis (EAE) through CD4+ T regulatory cell induction [11•].

Control of immunity by DCs is no doubt affected by costimulation and hence by the external factors that affect costimulatory molecule expression. Systemic administration of IL-4 inhibits both spontaneous and virus-induced diabetes. IL-4 prevents diabetes in non-obese diabetic mice by generating a Th2 response in a typically pathogenic Th1 environment [34]. The specific mechanism by which IL-4 inhibits lymphocytic choriomeningitis virus (LCMV)-induced diabetes was recently explored. King et al. [35^{••}] found that IL-4, selectively expressed in β cells, acts directly on local DCs by differentially altering expression of B7 molecules. Exposure of DCs to IL-4 caused CD80 levels to decrease and CD86 levels to increase, a combination that suppressed the effector function of pathogenic CD8+ T cells. Thus, costimulatory molecule engagements at the DC-T cell interface counter-regulate the effector function of autoreactive T cells. Similarly, blockade of CD40L protects mice from several autoimmune diseases. In an attempt to elucidate the mechanism by which anti-CD40L treatment inhibits virus-induced diabetes, Homann et al. [36^{••}] characterized a novel population of DCs that expanded in the spleens of LCMV-infected mice following anti-CD40L treatment. This unique cell population, which also displays features of natural killer (NK) cells and macrophages, completely inhibited diabetes upon transfer. Although the specific role of these regulatory DCs has not been resolved, it is intriguing to speculate that they counterregulate the function of pathogenic CD8+ T cells because they express high levels of B7-2 and low levels of B7-1.

Dendritic cell physiology in autoimmune tissues

Our understanding of the role of DCs in autoimmune disease stems, in part, from direct examination of these APCs in tissues of autoimmune subjects. DCs have been detected in lesions associated with numerous autoimmune diseases, including diabetes, rheumatoid arthritis (RA), psoriasis, EAE, thyroiditis, Sjogren's syndrome and SLE, and they are among the first cells to infiltrate target organs [37–44]. Unique functions of DCs in autoimmune tissues may coordinate the recruitment and/or activation of other immune players [8,45]. For example, abnormal chemokine secretion by DCs in tissues of nephritic mice preferentially acts on B1 cells, which may trigger autoantibody production in lupus [46•].

The origin of DCs in autoimmune lesions may illuminate their specific role(s) in autoimmune pathology. Although DCs in autoimmune lesions are primarily mature, it is unclear whether they arrive as fully mature cells or as DC precursors that undergo differentiation upon contact with specific tissue factors [8,38,39,47]. Santiago-Schwarz et al. [48] explored the composition of DCs in autoimmune synovial fluid and found that multiple stages of DC development were represented. In addition to mature DCs, they detected proliferating CD34-CD33+ myeloid progenitors that gave rise to non-proliferating myelodendritic progenitors. Interestingly, RA synovial fluid promoted differentiation of myelodendritic progenitors into functionally mature DCs in vitro; however, this was not the case when myelodendritic progenitors were cultured with synovial fluid from osteoarthritic patients or with normal human serum, or when CD34+ progenitors were cultured with RA synovial fluid. Thus, autoimmune synovial fluid contains factors that attract myelodendritic progenitors and promote their differentiation into mature DCs.

Likewise, Blanco et al. [49•] demonstrated that serum from SLE patients enhanced monocyte differentiation into immunostimulatory DCs and that this potential correlated directly with disease activity. IFN- α , which is elevated in the blood of patients with SLE, RA, Sjogren's syndrome and scleroderma, was the culprit driving DC development from monocytes. Paradoxically, the overall numbers of myeloid DCs, their monocyte precursors, and IFN-aproducing plasmacytoid DCs in patient blood were reduced. A decrease in blood DCs could be accounted for by augmented migration to tissues, as seen in murine lupus [46°,50]. Abnormalities in myeloid DC development and maturation have also been observed in murine models of diabetes and lupus [51–56]. Unfortunately, the reports are not in agreement with each other, preventing a general consensus from being reached. Elucidating these putative defects in myelopoiesis will be critical for understanding DC differentiation in autoimmune disease.

Conclusions

Our understanding of DCs and their roles in autoimmune disease has broadened in several ways this past year. We have learned that DCs not only promote immunity but also mediate peripheral T cell tolerance by direct elimination, T regulatory cell induction or counter-regulation. Tolerance can be induced by adoptive transfer of immature or semi-mature DCs, or by DCs presenting self-antigens under steady state conditions; however, the functional attributes that distinguish a tolerogenic from an 'autoimmunogenic' DC have yet to be defined. The efficacy of DCs in inducing peripheral tolerance may be useful for the treatment of ongoing human autoimmune conditions and preventing autoimmune disease onset in disease-prone subjects.

Studies of subjects with acute autoimmune disease indicate that DCs in autoimmune lesions exhibit an altered mature phenotype, whereas DCs obtained directly from blood or differentiated from blood precursors exhibit developmental defects as well as an altered immature phenotype; however, whether these alterations are causes or effects of autoimmune disease remains unclear. Taken together, these studies demonstrate that DCs are extremely versatile APCs capable of actively subduing autoimmunity under homeostatic or immunosuppressive conditions and mounting protective immunity during infection or inflammation. Autoimmune disease may arise when peripheral tolerance mechanisms are disrupted or when the immunogenicity of DCs is aimed at self-antigens.

Update

A specific role has been identified for the enigmatic CD4⁺ DC population in controlling autoimmune disease. In contrast to their CD8⁺ counterparts, which trigger autoimmunity via robust IL-12 production, splenic CD4⁺ DCs reverse CNS homogenate-induced EAE [57••]. CD4⁺ DCs that have internalized aggregated Ig-MOG via FcγRI suppress autoimmunity by secreting IL-10.

Acknowledgements

I would like to thank Ananda Goldrath, Torben Lund, and Diane Mathis for critical assessment of the manuscript and stimulating discussions.

References and recommended reading

Papers of particular interest, published within the annual period of review, have been highlighted as:

- of special interest
- · of outstanding interest
- Banchereau J, Briere F, Caux C, Davoust J, Lebecque S, Liu Y-J, Pulendran B, Palucka K: Immunobiology of dendritic cells. Annu Rev Immunol 2000, 18:767-811.
- Steinman RM, Nussenzweig MC: Avoiding horror autotoxicus: the importance of dendritic cells in peripheral T cell tolerance. Proc Natl Acad Sci USA 2002, 99:351-358.
- Knight SC, Mertin J, Stackpoole A, Clark J: Induction of immune responses in vivo with small numbers of veiled (dendritic) cells. Proc Natl Acad Sci USA 1983, 80:6032-6035.
- Knight SC, Farrant J, Chan J, Bryant A, Bedford PA, Bateman C: Induction of autoimmunity with dendritic cells: studies on thyroiditis in mice. *Clin Immunol Immunopathol* 1988, 48:277-289.
- Dittel BN, Visintin I, Merchant RM, Janeway CA Jr: Presentation of the self antigen myelin basic protein by dendritic cells leads to experimental autoimmune encephalomyelitis. *J Immunol* 1999, 163:32-39.
- Weir CR, Nicolson K, Backstrom BT: Experimental autoimmune encephalomyelitis induction in naive mice by dendritic cells presenting a self-peptide. *Immunol Cell Biol* 2002, 80:14-20.
- Ludewig B, Odermatt B, Landmann S, Hengartner H, Zinkernagel RM: Dendritic cells induce autoimmune diabetes and maintain disease via *de novo* formation of local lymphoid tissue. *J Exp Med* 1998, 188:1493-1501.
- 8. Green EA, Eynon EE, Flavell RA: Local expression of TNFalpha in neonatal NOD mice promotes diabetes by enhancing presentation of islet antigens. *Immunity* 1998, **9**:733-743.

- Mehling A, Loser K, Varga G, Metze D, Luger TA, Schwarz T, Grabbe S, Beissert S: Overexpression of CD40 ligand in murine epidermis results in chronic skin inflammation and systemic autoimmunity. J Exp Med 2001, 194:615-628.
- Clare-Salzler MJ, Brooks J, Chai A, Van Herle K, Anderson C: Prevention of diabetes in nonobese diabetic mice by dendritic cell transfer. J Clin Invest 1992, 90:741-748.
- Menges M, Rossner S, Voigtlander C, Schindler H, Kukutsch NA,
 Bogdan C, Erb K, Schuler G, Lutz MB: Repetitive injections of dendritic cells matured with tumor necrosis factor alpha induce antigen-specific protection of mice from autoimmunity. *J Exp Med* 2002. 195:15-21.

This study underscores the important point that not all DC maturation stimuli should be regarded as equal. The authors demonstrate that serial transfers of bone marrow-derived DCs pretreated with MOG-peptide and TNF- α completely protect recipients from EAE by inducing CD4+T regulatory cells, whereas anti-CD40 and LPS treatment generated classical mature DCs with no tolerogenic potential. Thus, partially mature or intermediate DCs are conditioned, albeit in unknown ways, for peripheral tolerance induction.

- Shinomiya M, Fazle Akbar SM, Shinomiya H, Onji M: Transfer of dendritic cells (DC) ex vivo stimulated with interferon- gamma (IFN-gamma) down-modulates autoimmune diabetes in nonobese diabetic (NOD) mice. Clin Exp Immunol 1999, 117:38-43.
- Naumov YN, Bahjat KS, Gausling R, Abraham R, Exley MA, Koezuka Y, Balk SB, Strominger JL, Clare-Salzer M, Wilson SB: Activation of CD1d-restricted T cells protects NOD mice from developing diabetes by regulating dendritic cell subsets. Proc Natl Acad Sci USA 2001, 98:13838-13843.
- Feili-Hariri M, Dong X, Alber SM, Watkins SC, Salter RD, Morel PA: Immunotherapy of NOD mice with bone marrow-derived dendritic cells. Diabetes 1999, 48:2300-2308.
- Feili-Hariri M, Falkner DH, Morel PA: Regulatory Th2 response induced following adoptive transfer of dendritic cells in prediabetic NOD mice. *Eur J Immunol* 2002, 32:2021-2030.
- Pugh CW, MacPherson GG, Steer HW: Characterization of nonlymphoid cells derived from rat peripheral lymph. J Exp Med 1983, 157:1758-1779.
- Huang F-P, Platt N, Wykes M, Major JR, Powell TJ, Jenkins CD, MacPherson GG: A discrete subpopulation of dendritic cells transports apoptotic intestinal epithelial cells to T cell areas of mesenteric lymph nodes. J Exp Med 2000, 191:435-443.
- Hemmi H, Yoshino M, Yamazaki H, Naito M, Iyoda T, Omatsu Y,
 Shimoyama S, Letterio JJ, Nakabayashi T, Tagaya H *et al.*: Skin antigens in the steady state are trafficked to regional lymph nodes by transforming growth factor-beta1-dependent cells. *Int Immunol* 2001, 13:695-704.

Using hyperpigmented mice to monitor trafficking of skin-restricted antigens, the authors show that melanin granules are transported to draining LNs in the absence of inflammation. Melanin granules are undetectable in LNs of transforming growth factor (TGF)- β -deficient mice (which lack epidermal DCs or Langerhans cells), indicating that insoluble skin antigens are continuously ferried to LNs by migratory DCs that have captured melanocyte fragments rather than by lymph- or macrophage-mediated transport pathways.

- 19. Hawiger D, Inaba K, Dorsett Y, Guo M, Mahnke K, Rivera M,
- Ravetch JV, Steinman RM, Nussenzweig MC: Dendritic cells induce peripheral T cell unresponsiveness under steady state conditions in vivo. J Exp Med 2001, 194:769-779.

This work establishes a functional role for DC-mediated antigen transport that occurs under homeostatic conditions. A monoclonal antibody specific for DEC205, an endocytic receptor highly expressed on DCs, was used to efficiently target specific peptides to DCs in *vivo*. The authors show that presentation of soluble antigen by DCs in healthy, uninfected subjects leads to tolerance by deletion of antigen-specific CD4⁺ T cells.

- Kurts C, Kosaka H, Carbone FR, Miller JFAP, Heath WR: Class Irestricted cross-presentation of exogenous self antigens leads to deletion of autoreactive CD8+ T cells. J Exp Med 1997, 186:239-245.
- Albert ML, Jegathesan M, Darnell RB: Dendritic cell maturation is required for the cross-tolerization of CD8+ T cells. Nat Immunol 2001, 2:1010-1017.

This study characterizes the requirements for *in vitro* cross tolerisation of human memory CD8⁺ T cells. The authors demonstrate that an increase in B7 expression induced by CD40 engagement is necessary for deletion of antigen-specific CD8⁺ T cells by DCs.

 Jansen A, van Hagen M, Drexhage HA: Defective maturation and function of antigen-presenting cells in type 1 diabetes. *Lancet* 1995, 345:491-492.

- Tas M, Haan-Meulman M, Kabel PJ, Drexhage HA: Defects in monocyte polarization and dendritic cell clustering in patients with Graves' disease. A putative role for a non-specific immunoregulatory factor related to retroviral p15E. Clin Endocrinol (Oxt) 1991, 34:441-448.
- Takahashi K, Honeyman MC, Harrison LC: Impaired yield, phenotype, and function of monocyte-derived dendritic cells in humans at risk for insulin-dependent diabetes. *J Immunol* 1998, 161:2629-2635.
- Scheinecker C, Zwolfer B, Koller M, Manner G, Smolen JS: Alterations of dendritic cells in systemic lupus erythematosus: phenotypic and functional deficiencies. *Arthritis Rheum* 2001, 44:856-865.
- Huang YM, Stoyanova N, Jin YP, Teleshova N, Hussien Y, Xiao BG, Fredrikson S, Link H: Altered phenotype and function of blood dendritic cells in multiple sclerosis are modulated by IFN-beta and IL-10. *Clin Exp Immunol* 2001, 124:306-314.
- 27. Pugliese A, Brown D, Garza D, Murchison D, Zeller M, Redondo M,
- Diez J, Eisenbarth GS, Patel DD, Ricordi C: Self-antigen-presenting cells expressing diabetes-associated autoantigens exist in both thymus and peripheral lymphoid organs. J Clin Invest 2001, 107:555-564.

This investigation demonstrates that DCs in human spleen and thymus express tissue-specific autoantigens such as GAD, proinsulin and IA-2. This evidence supports the provocative hypothesis that a specialized subset of extrathymic DCs induces tolerance by presenting endogenously expressed autoantigens followed by direct killing of autoreactive T lymphocytes.

- Roncarolo M-G, Levings MK, Traversari C: Differentiation of T regulatory cells by immature dendritic cells. J Exp Med 2001, 193:F5-F9.
- Jonuleit H, Schmitt E, Schuler G, Knop J, Enk AH: Induction of interleukin 10-producing, nonproliferating CD4(+) T cells with regulatory properties by repetitive stimulation with allogeneic immature human dendritic cells. J Exp Med 2000, 192:1213-1222.
- 30. Dhodapkar MV, Steinman RM, Krasovsky J, Munz C, Bhardwaj N:
 Antigen-specific inhibition of effector T cell function in humans after injection of immature dendritic cells. J Exp Med 2001, 193:233-238.

This study establishes that treatment of humans with antigen-bearing immature DCs suppresses cognate CD8⁺ T cell function by inducing regulatory T cells within one week of injection. In a follow-up study (see [31••]), the authors demonstrate that CD8⁺ T cell effector function bounced back within six months of the initial DC injection. Regulatory T cells generated during the first week after immunization were capable of suppressing the functions of restored CD8⁺ effectors.

- Dhodapkar MV, Steinman RM: Antigen-bearing immature dendritic
 cells induce peptide-specific CD8(+) regulatory T cells *in vivo* in humans. *Blood* 2002, 100:174-177.
- See annotation to [30 ••]
- Steinman RM, Turley S, Mellman I, Inaba K: The induction of tolerance by dendritic cells that have captured apoptotic cells. *J Exp Med* 2000, 191:411-416.
- Hugues S, Mougneau E, Ferlin W, Jeske D, Hofman P, Homann D,
 Beaudoin L, Schrike C, Von Herrath M, Lehuen A, et al.: Tolerance to islet antigens and prevention from diabetes induced by limited apoptosis of pancreatic beta cells. *Immunity* 2002, 16:169-181.

This work demonstrates that deliberate induction of pancreatic β cell apoptosis protects mice from adoptively transferred diabetes by inducing regulatory T cells. The evidence provided here suggests that low grade apoptosis releases β cell antigen for presentation by DCs after migration to pancreatic LNs and conditions the DC for tolerance induction.

- Cameron MJ, Arreaza GA, Zucker P, Chensue SW, Strieter RM, Chakrabarti S, Delovitch TL: IL-4 prevents insulitis and insulindependent diabetes mellitus in nonobese diabetic mice by potentiation of regulatory T helper-2 cell function. *J Immunol* 1997, 159:4686-4692.
- 35. King C, Mueller HR, Malo CM, Murali-Krishna K, Ahmed R, King E,
 Sarvetnick N: Interleukin-4 acts at the locus of the antigenpresenting dendritic cell to counter-regulate cytotoxic CD8+ T-cell responses. Nat Med 2001, 7:206-214.

This study defines the mechanism by which islet-restricted expression of IL-4 inhibits LCMV-induced diabetes but not insulitis. The authors demonstrate that IL-4 skewed the functional capacity of local DCs by increasing their expression of B7-2 and reducing their expression of B7-1. Engagement of islet-specific CD8⁺ T cells by IL-4-exposed DCs impaired their differentiation into functional CTL.

 ξ36. Homann D, Jahreis A, Wolfe T, Hughes A, Coon B, van Stipdonk MJ,
 Prilliman KR, Schoenberger SP, Von Herrath MG: CD40L blockade prevents autoimmune diabetes by induction of bitypic NK/DC regulatory cells. *Immunity* 2002, 16:403-415.

This study characterizes a novel population of DX5⁺ regulatory DCs that inhibit LCMV-induced diabetes in response to CD40L blockade. This cell population exhibits functional and phenotypic features of NK cells and macrophages; however, their origin and mechanism of action remain unclear.

- Jansen A, Homo-Delarche F, Hooijkaas H, Leenen PJ, Dardenne M, Drexhage HA: Immunohistochemical characterization of monocytes-macrophages and dendritic cells involved in the initiation of the insulitis and beta-cell destruction in NOD mice. Diabetes 1994, 43:667-675.
- Thomas R, Davis LS, Lipsky PE: Rheumatoid synovium is enriched in mature antigen-presenting dendritic cells. J Immunol 1994, 152:2613-2623.
- Demidem A, Taylor JR, Grammer SF, Streilein JW: T-lymphocyteactivating properties of epidermal antigen-presenting cells from normal and psoriatic skin: evidence that psoriatic epidermal antigen-presenting cells resemble cultured normal Langerhans cells. J Invest Dermatol 1991, 97:454-460.
- Serafini B, Columba-Cabezas S, Di Rosa F, Aloisi F: Intracerebral recruitment and maturation of dendritic cells in the onset and progression of experimental autoimmune encephalomyelitis. *Am J Pathol* 2000, **157**:1991-2002.
- Voorby HA, Kabel PJ, de Haan M, Jeucken PH, van der Gaag RD, de Baets MH, Drexhage HA: Dendritic cells and class II MHC expression on thyrocytes during the autoimmune thyroid disease of the BB rat. Clin Immunol Immunopathol 1990, 55:9-22.
- Quadbeck B, Eckstein AK, Tews S, Walz M, Hoermann R, Mann K, Gieseler R: Maturation of thyroidal dendritic cells in Graves' disease. Scand J Immunol 2002, 55:612-620.
- 43. Ozaki Y, Amakawa R, Ito T, Iwai H, Tajima K, Uehira K, Kagawa H, Uemura Y, Yamashita T, Fukuhara S: Alteration of peripheral blood dendritic cells in patients with primary Sjogren's syndrome. *Arthritis Rheum* 2001, 44:419-431.
- Denfeld RW, Kind P, Sontheimer RD, Schopf E, Simon JC: In situ expression of B7 and CD28 receptor families in skin lesions of patients with lupus erythematosus. Arthritis Rheum 1997, 40:814-821.
- Dahlen E, Dawe K, Ohlsson L, Hedlund G: Dendritic cells and macrophages are the first and major producers of TNF-alpha in pancreatic islets in the nonobese diabetic mouse. *J Immunol* 1998, 160:3585-3593.
- 46. Ishikawa S, Sato T, Abe M, Nagai S, Onai N, Yoneyama H, Zhang Y,
 Suzuki T, Hashimoto S, Shirai T *et al.*: Aberrant high expression of B lymphocyte chemokine (BLC/CXCL13) by C11b+CD11c+ dendritic cells in murine lupus and preferential chemotaxis of B1 cells towards BLC. J Exp Med 2001, 193:1393-1402.

This work demonstrates that thymic DCs in mice with lupus produce abnormally high levels of B lymphocyte chemokine and that CXCR5^{hi} B1 cells are selectively recruited by this chemokine. Evidence provided in this report supports the hypothesis that abnormal chemokine production by DCs coordinates the formation of autoimmune infiltrates by attracting specific lymphocyte populations.

- Pettit AR, MacDonald KP, O'Sullivan B, Thomas R: Differentiated dendritic cells expressing nuclear RelB are predominantly located in rheumatoid synovial tissue perivascular mononuclear cell aggregates. Arthritis Rheum 2000, 43:791-800.
- Santiago-Schwarz F, Anand P, Liu S, Carsons SE: Dendritic cells (DCs) in rheumatoid arthritis (RA): progenitor cells and soluble factors contained in RA synovial fluid yield a subset of myeloid DCs that preferentially activate Th1 inflammatory-type responses. *J Immunol* 2001, 167:1758-1768.
- 49. Blanco P, Palucka AK, Gill M, Pascual V, Banchereau J: Induction of
 dendritic cell differentiation by IFN-alpha in systemic lupus erythematosus. *Science* 2001, 294:1540-1543.

This paper defines unique functional attributes of blood myeloid cells and serum components in pediatric SLE patients that might play important roles in autoimmune disease.

 Kalled SL, Cutler AH, Burkly LC: Apoptosis and altered dendritic cell homeostasis in lupus nephritis are limited by anti-CD154 treatment. J Immunol 2001, 167:1740-1747.

- Weaver DJ Jr., Poligone B, Bui T, Abdel-Motal UM, Baldwin AS Jr., Tisch R: Dendritic cells from nonobese diabetic mice exhibit a defect in NF- kappa B regulation due to a hyperactive I kappa B kinase. J Immunol 2001, 167:1461-1468.
- Feili-Hariri M, Morel PA: Phenotypic and functional characteristics of BM-derived DC from NOD and non-diabetes-prone strains. *Clin Immunol* 2001, 98:133-142.
- Strid J, Lopes L, Marcinkiewicz J, Petrovska L, Nowak B, Chain BM, Lund T: A defect in bone marrow derived dendritic cell maturation in the nonobese-diabetic mouse. *Clin Exp Immunol* 2001, 123:375-381.
- Prasad SJ, Goodnow CC: Cell-intrinsic effects of non-MHC NOD genes on dendritic cell generation *in vivo*. Int Immunol 2002, 14:677-684.
- 55. Prasad SJ, Goodnow CC: Intrinsic *in vitro* abnormalities in dendritic cell generation caused by non-MHC non-obese diabetic genes. *Immunol Cell Biol* 2002, **80**:198-206.
- 56. Steptoe RJ, Ritchie JM, Harrison LC: Increased generation of dendritic cells from myeloid progenitors in autoimmune-prone nonobese diabetic mice. *J Immunol* 2002, **168**:5032-5041.
- 57. Legge KL, Greg RK, Maldonado-Lopez R, Li L, Caprio JC, Moser M,
 Zaghouani H: On the role of dendritic cells in peripheral T cell tolerance and modulation of autoimmunity. *J Exp Med* 2002,

196:217-227. Using adoptive transfers of specific DC populations, the authors discover a function for CD4⁺ DCs in establishing peripheral tolerance. IL-10 production by CD4⁺ DCs counters the effects of IL-12 made by CD8 α ⁺ DCs, leading to reversal of ongoing EAE.